

Investigation of infection

Investigation of infection

❖ The aims of investigating a patient with suspected infection are :-

- Confirm the presence of infection.
- Identify the specific pathogen(s).
- Identify its susceptibility to specific antimicrobial agents in order to optimize therapy.

❖ The presence of infection may be suggested by ;

- Identifying proteins that are produced in response to pathogens as part of the innate immune.
- Acute phase responses.

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❖ Pathogens may be detected :-

- Directly (e.g. by culturing a normally sterile body site).
- Indirect detection by identifying the host response to the organism.

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❖ Tests used to diagnose infection:-

➤ **Non-specific markers of inflammation/infection :- e.g.**

✓ **WCC.**

✓ **CRP.**

✓ **Procalcitonin.**

✓ **Serum lactate.**

✓ **Cell counts in urine or(CSF).**

✓ **CSF protein and glucose.**

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❖ Tests used to diagnose infection:-

➤ Direct detection of organisms or organism components :-

- ✓ Microscopy.

- ✓ Detection of organism components (e.g. antigen, toxin).

- ✓ Nucleic acid amplification (e.g. polymerase chain reaction).

➤ Culture of organisms :- ± Antimicrobial susceptibility testing.

➤ Tests of the host's specific immune response :-

- ✓ Antibody detection.

- ✓ Interferon-gamma release assays (IGRA).

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❖ How to provide samples for microbiological samplings :-

➤ Communicate with the laboratory

✓ Discuss with laboratory staff before collection.

✓ Communication is key to optimizing microbiological diagnosis.

✓ Discuss with laboratory staff before hand than to risk diagnostic delay by inappropriate sampling or sample handling.

➤ Take samples based on a clinical diagnosis

✓ Sampling in the absence of clinical evidence of infection is rarely appropriate.

➤ Use the correct container

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❖ How to provide samples for microbiological samplings :-

➤ Use the correct container

✓ Certain tests require proprietary sample collection equipment.

➤ Follow sample collection procedures

✓ Failure to follow sample collection instructions can result in

○ False-positive (e.g. contamination of blood culture samples).

○ False-negative (e.g. collection of insufficient blood for culture) results.

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❖ How to provide samples for microbiological samplings :-

➤ **Label sample and request form correctly according to local policies,**

➤ **Use appropriate packaging**

✓ **Close sample containers tightly and package securely (usually in sealed plastic bags).**

✓ **Attach request forms to samples but not in the same compartment.**

➤ **Manage storage and transport**

✓ **Transport samples to the microbiology laboratory quickly.**

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❖ Direct detection of pathogens

- Provide rapid results and enable detection of organisms that cannot be grown easily on artificial culture media, such as Chlamydia spp.
- Provide information on antimicrobial sensitivity, e.g. Mycobacterium Tuberculosis.

❖ Detection of whole organisms

- Whole organisms are detected by examination of biological fluids or tissue using a microscope.

❑ Bright field microscopy.

❑ Dark field microscopy.

❑ Electron microscopy.

❑ Flow cytometry.

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❖ Detection of components of organisms

➤ Components of microorganisms detected for diagnostic purposes include;-

✓ Nucleic acids.

✓ Cell wall molecules.

✓ Toxins.

✓ Other antigens.

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❖ Detection of components of organisms

✓ Nucleic acid amplification tests

- Specific sequences of microbial DNA and RNA are identified using a nucleic acid primer.
- The most commonly used amplification method is the polymerase chain reaction (PCR).
- Reverse transcription (RT) PCR is used to detect RNA from RNA viruses (e.g. hepatitis C virus and HIV-1).
- In multiplex PCR, multiple primer pairs are used to enable detection of several different organisms at once.
- NAATs are the most sensitive direct detection methods and are also relatively rapid.

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❖ Detection of components of organisms

✓ Nucleic acid amplification tests

- Used widely in virology.
- In bacteriology, PCR is used to examine CSF, blood, tissue and genital samples, and multiplex PCR is being developed for use in faeces.
- Helpful for microorganisms that cannot be readily cultured.
- Increasingly used in mycology and parasitology.

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- ❖ Detection of components of organisms
- ✓ Nucleic acid amplification tests

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❖ Culture :-

- Microorganisms may be detected and further characterized by culture from clinical samples.
- In vitro culture of bacteria and fungi is used to;
- ✓ Confirm the presence of pathogens.
- ✓ Allow identification.
- ✓ Test antimicrobial susceptibility.
- ✓ Subtype the organism for epidemiological purposes.
- **limitations:** results are not immediate, even for organisms that are easy to grow, and negative cultures rarely exclude infection.

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❖ Culture :-

- Organisms such as *Mycobacterium tuberculosis* are slow-growing, typically taking at least 2 weeks, Even in rapid-culture systems.
- *Mycobacterium leprae* and *Tropheryma whipplei*, cannot be cultivated on artificial media.
- Others (e.g. *Chlamydia* spp. and viruses) grow only in culture systems, which are slow and labor-intensive.
- The terms 'bacteremia' and 'fungaemia' describe the presence of bacteria and fungi in the blood.
- 'Blood-stream infection' is the association of bacteremia/fungaemia with clinical evidence of infection.

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❖ Culture :-

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❖ Indirect detection of pathogens :-

- Used to detect the host's immune (antibody) response to a specific microorganism.
- Can enable the diagnosis of infection with organisms that are difficult to detect by other methods or are no longer present in the host.
- The term 'serology' describes tests carried out on serum and includes both antigen (direct) and antibody (indirect) detection.
- Organism-specific antibody detection is applied mainly to blood.
- Results are typically expressed as titers.

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❖ Indirect detection of pathogens :-

- 'Seroconversion' is defined as either a change from negative to positive detection or a fourfold rise in titer between acute and convalescent serum samples.
- An acute sample is usually taken during the first week of disease and the convalescent sample 2–4 weeks later.
- Earlier diagnosis can be achieved by detection of immunoglobulin M (IgM) antibodies, which are produced early in infection.
- A limitation of these tests is that antibody production requires a fully functional host immune system, so there may be false-negative results in immunocompromised patients.

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❖ Indirect detection of pathogens :-

❑ Enzyme-linked immunosorbent assay

- The principles of the (ELISA, EIA) are in assays rely on linking an antibody with an enzyme that generates a color change on exposure to a chromogenic substrate.
- Configurations allow detection of antigens or specific subclasses of immunoglobulin (e.g. IgG, IgM, IgA).
- ELISA may also be adapted to detect PCR products.

❑ Immunoblot (Western blot)

- Microbial proteins are separated and transferred (blotted) on to a nitrocellulose membrane, which is incubated with patient serum.
- Binding of specific antibody is detected with an enzyme–anti-immunoglobulin.
- A highly specific test, which may be used to confirm the results of less specific tests.

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❖ Indirect detection of pathogens :-

❑ Immunofluorescence assays

- Indirect immunofluorescence assays (IFAs) detect antibodies.
- Any virus-specific antibody present in the serum binds to antigen.
- Fluorescence is visualized using a microscope.
- This method can also detect organisms in clinical samples, using a specific antibody.

❑ Complement fixation test

- Any specific antibody in the serum will complex with the antigen.
- The degree of erythrocyte lysis reflects the remaining complement and is inversely proportional to the quantity of the specific antigen–antibody complex present.

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❖ Indirect detection of pathogens :-

☐ Agglutination tests

- When antigens are present on the surface of particles and cross-linked with antibodies, visible clumping (or 'agglutination') occurs.
- The test lacks sensitivity and specificity but is still used to diagnose rickettsial infection in resource-limited settings.
- The Widal test reaction uses a suspension of *Salmonella typhi* and *S. Paratyphi* 'A' and 'B', treated to retain only 'O' and 'H' antigens.
- Antigens are kept to detect corresponding antibodies in serum from a patient suspected of having typhoid fever.
- The test is not specific but is still used in some parts of the world.

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❖ Indirect detection of pathogens :-

☐ Antibody-independent specific immunological tests

- The interferon-gamma release assay (IGRA) is being used increasingly to diagnose (LTBI).
- IGRA cannot distinguish between latent and active tuberculosis infection.
- Use only in countries where the background incidence of tuberculosis is low.

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❖ Antimicrobial susceptibility testing :-

- If growth of microorganisms in culture is inhibited by the addition of an antimicrobial agent, the organism is considered to be susceptible to that antimicrobial.
- Bacteriostatic agents cause reversible inhibition of growth.
- Bactericidal agents cause cell death.
- Fungistatic/fungicidal are equivalent for antifungal agents.
- Virustatic/virucidal for antiviral agents.

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❖ Antimicrobial susceptibility testing :-

- The lowest concentration of antimicrobial agent at which growth is inhibited is the minimum inhibitory concentration (MIC)
- The lowest concentration that causes cell death is the minimum bactericidal concentration (MBC).
- If the MIC is less than or equal to a predetermined breakpoint threshold, the organism is considered susceptible.
- If the MIC is greater than the breakpoint, it is resistant.

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❖ Antimicrobial susceptibility testing :-

- The relationship between in vitro antimicrobial susceptibility and clinical response is depends on;
- ✓ Immune status.
- ✓ Pharmacokinetic variability.
- ✓ Comorbidities that may influence pharmacokinetics or pharmacodynamics.
- ✓ Antibiotic dosing.
- ✓ MIC/MBC.

GOOD LUCK